

AMBIENT TEMPERATURE SHIPPING AND STORAGE FOR MOLECULAR INFECTIOUS DISEASE TESTING: ViveST®

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Introduction

A key component of molecular infectious disease testing centers around the preservation of biological samples during shipping and storage. Typically samples are collected at one site and shipped to a central laboratory for testing requiring careful temperature control and special packaging to minimize effects of harsh shipping conditions. Time elapses between sample collection and analysis; therefore, it is imperative to have a shipping and storage process that is efficient and preserves sample integrity over time.

The current standards require dry ice, special packaging, and ultra-cold storage, all of which is expensive and laborious. This study evaluated the stability of HIV-1 plasma samples shipped and stored on ViveST®, a transformational dried ambient temperature storage and transportation device (ViveBio LLC, Alpharetta, GA, USA).

Methods

To evaluate effects of shipping, 4 sets of HIV-1 plasma samples were prepared, loaded onto ViveST, dried overnight at room temperature (RT), and capped. Each set contained twenty 1.15 mL aliquots of HIV-1 plasma (4 levels, 5 replicates each) and 1 negative control.

1 set was stored frozen (-80°C) and 3 sets were each packaged in standard cardboard boxes and shipped via FedEx from bioMONTR Labs in North Carolina to 1 of 3 distinct geographic locations across the continental United States (Northeast, Southwest and West Coast). The 3 sets were returned (via FedEx) to bioMONTR Labs for analysis.

All samples were recovered using 1.15 mL of molecular grade water and analyzed with Abbott's RealTime HIV-1 assay (0.6 mL protocol, Abbott Laboratories, Illinois, USA).

One-Way Omnibus ANOVA in R was used to evaluate the variance between replicates shipped to 3 sites.

To evaluate stability of HIV-1 plasma stored on the ViveST device, 21 sets of HIV-1 plasma were prepared, loaded onto ViveST, dried overnight and stored for up to 2 months. Each set contained twenty 1.1 mL aliquots of HIV-1 plasma (4 levels, 5 replicates each) and 1 negative control.

Seven sets were stored at each of the three different conditions (RT, 4°C, and 40°C/75% RH). One set was removed from each storage condition at days 1, 3, 7, 10, 14, 21, and 62 and analyzed.

All samples were recovered using 1.1 mL of molecular grade water and analyzed with Abbott's RealTime HIV-1 assay (0.6 mL protocol, Abbott Laboratories, Illinois, USA). For comparative purposes, identical 1.1 mL aliquots (4 levels, 5 replicates each) of frozen plasma were analyzed.

Linear Regression Analysis was performed to evaluate effects of storage at different conditions over time.

Results

For HIV-1 plasma shipped on ViveST, the average reduction in viral load when compared to frozen plasma was 1.07 LOG c/mL (See Table 1). A linear fit ($R^2 > 0.99$) was retained across all levels/all sites (See Figure 1).

The omnibus ANOVA test results indicate NO significant differences between the measurements shipped to the 3 sites (P-Value = 0.9897). Box Plot of viral load results demonstrate that the data from the three sites are equivalent (See Figure 2).

For HIV-1 plasma stored on ViveST for a 62-day period (ambient temperature), the maximum reduction recorded when compared to frozen plasma (data not shown) was 0.91 LOG c/mL, 0.84 LOG c/mL (4°C), and 1.69 LOG c/mL (40°C/75%RH).

A linear fit ($R^2 > 0.98$) was retained over the course of the 62-day study as indicated by linear regression analysis (See Figure 3, ambient storage only).

The data demonstrated that samples stored on ViveST over a 62-day period at RT, 4°C, and 40°C/75% RH yielded reproducible results. (See Figure 4).

Table 1. Summary of Abbott RealTime HIV-1 Assay Data_ViveST Shipping Study

Level	Replicate	Frozen	Shipping - Ambient Conditions		
		Log c/mL	Site 1	Site 2	Site 3
1	A	6.05	4.84	4.81	4.89
	B	5.98	4.87	4.83	4.83
	C	5.95	4.82	4.84	4.86
	D	5.95	4.82	4.83	4.89
	E	6.03	4.90	4.80	4.88
	Mean	5.99	4.85	4.82	4.87
2	A	4.81	3.75	3.76	3.87
	B	4.93	3.73	3.68	3.78
	C	4.93	3.71	3.75	3.81
	D	4.93	3.69	3.72	3.84
	E	4.90	3.73	3.75	3.76
	Mean	4.90	3.72	3.73	3.81
3	A	3.78	2.88	2.86	2.96
	B	3.92	2.86	2.81	2.85
	C	3.86	2.88	2.88	2.9
	D	3.92	2.84	2.87	2.68
	E	3.88	2.82	2.95	2.85
	Mean	3.87	2.86	2.87	2.85
4	A	2.99	1.80	2.06	2.03
	B	2.91	1.73	2.13	2.01
	C	2.82	2.18	1.84	1.22
	D	2.95	1.84	1.99	2.13
	E	2.75	1.58	2.00	2.11
	Mean	2.88	1.83	2.00	1.90
Summary	Std deviation	0.10	0.22	0.11	0.38
	% CV	0.12	0.28	0.13	0.48

NOTE: Highlighted cells indicate samples where m2000r1 results were reported as <1.60 LOG c/ml (<40 c/mL). Results were manually calculated using the calibration curve.

Results Cont.

Figure 1. Linear Regression: ViveST Shipping Study

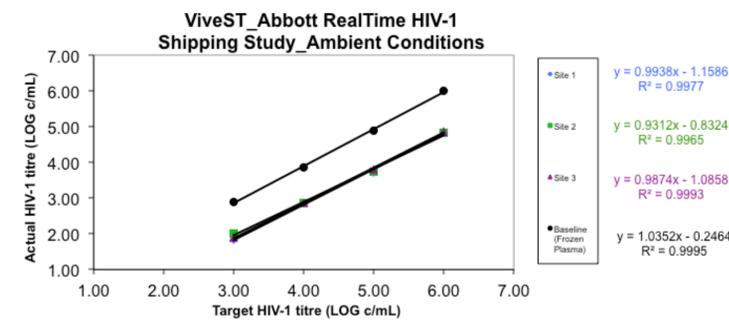


Figure 2. Boxplot: ViveST Shipping Study

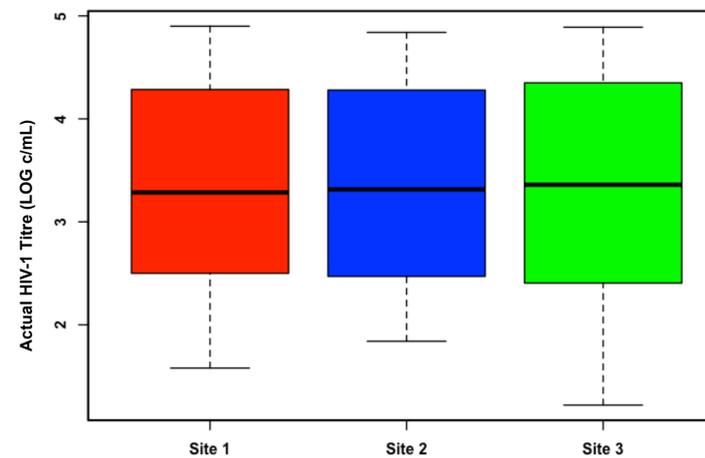
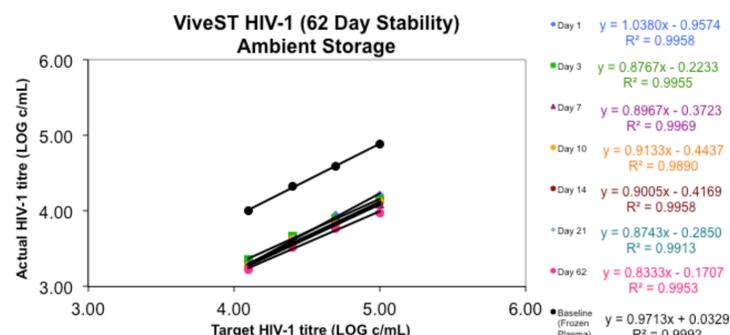
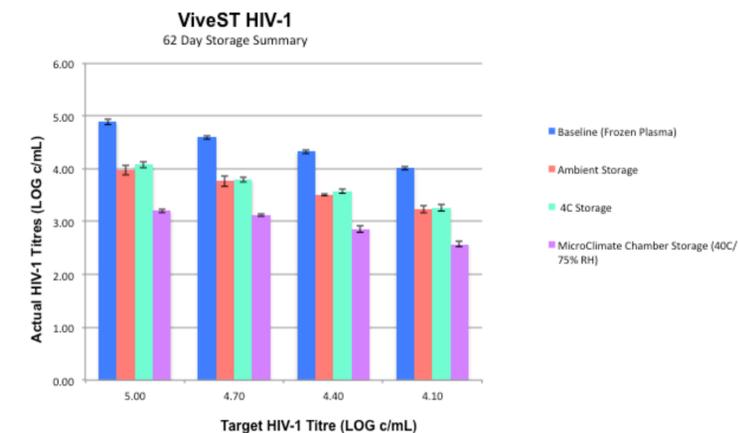


Figure 3. Linear Regression: ViveST Processed HIV-1 Plasma Ambient Storage through 62 Days



Results Cont.

Figure 4. ViveST Processed HIV-1 Plasma: Comparison Across All Storage Conditions (62 Days Storage)



Conclusions

- There is no significant difference between the HIV RNA levels of samples recovered from ViveST after shipping to 3 different sites. (P Value = 0.9897).
- The linear responses over time coupled with the high degree of precision and reproducibility observed with ViveST imply application of a conversion factor could be utilized to account for any reduction of viral RNA recovery to convert ViveST values to frozen values.
- ViveST provides significant cost savings as compared to dry ice shipments and can enhance access to healthcare globally while significantly reducing the cost burden associated with frozen samples.
- ViveST exhibits great potential for shipping and storing samples for viral load testing worldwide.

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